

## Imidazo[1,2-a]pyridines as HIV NNRTIs

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## Introduction

The implementation of highly active antiretroviral therapy (HAART) has dramatically altered both the life expectancy and quality of life of HIV-AIDS sufferers. Most first-line regimen drug cocktails are made up of a combination of nucleoside reverse transcriptase inhibitors (NRTIs) and non-nucleoside reverse transcriptase inhibitors (NNRTIs). These drugs inhibit reverse transcriptase (RT) by binding to a lipophilic, non-substrate binding pocket located about 10Å from the substrate binding site. Screening of a small inhouse library of compounds for activity against HIV-1 RT led to identification of imidazo[1,2-a]pyridines as potential NNRTIs. A broader synthesis and testing campaign was carried out based on this result.

## **Results and Discussion**

Compound 1, an imidazo[1,2-a]pyridine, was identified in the preliminary screen and served as the hit compound for this study. The Groebke multi-component reaction serves as a versatile method for the rapid assembly of imidazo[1,2-a]pyridines (Scheme 1). Several small libraries were generated by varying functionality on the aldehyde, isocyanide and 2-aminopyridine components. All compounds were tested against HIV-1 RT and selected compounds were tested in a cell-based anti-HIV assay (PBMC or MAGI).

$$\begin{array}{c} \begin{array}{c} R_1 \\ \\ \end{array} \\ \begin{array}{c} N \end{array} + \begin{array}{c} R_2 - \text{CHO} \end{array} + \begin{array}{c} R_2 - \text{CHO} \end{array} + \begin{array}{c} R_3 - \text{Res} \\ \\ \end{array} \\ \begin{array}{c} R_2 - \text{CHO} \end{array} \end{array}$$

A set of first-generation analogues incorporating maximum structural diversity in the isocyanide and aldehyde components was prepared and a selection of these is shown in Table 1.

Table 1. Selected first generation analogues (R1 = H)

No.	R <sub>2</sub>	R₃ Cyclohexyl 2-Pentyl	Biological Activity			
			% Residual Enzyme Activity†	IC <sub>50</sub> (μM) Enzymatic Assay	IC <sub>50</sub> (µM) PBMC <sup>§</sup> /MAGI <sup>¶</sup> Assay	
1	iso-Propyl		40 44	29 ND#	40 <sup>§</sup> 33¶	
2	2-Chlorophenyl					
3	*Q	Adamantyl	78	ND	ND	
4	2-Furyl	Cyclohexyl	71	ND	95§	
5	2-Furyl	a Dottse	36	ND	44¶	
6	2-Chlorophenyl	1-Pentyl 2-Naphthyl Cyclohexyl 2-Pentyl	55 100 87 55	ND ND ND ND	>200¶ ND 32 <sup>§</sup> >200¶	
7	4-Methoxyphenyl					
8	2-Hydroxyphenyl					
9	iso-Propyl					
10	2-Chlorophenyl	Cyclohexyl	7	4	0.16§ and 0.41¶	
11	4-Methoxyphenyl	+××	83	ND	ND	
12	Phenyl	Cyclohexyl	75	ND	ND	
13	iso-Propyl	1-Pentyl	84	ND	>200¶	
14	*	Cyclohexyl	25	14	102¶	
15	-}/-5	Cyclohexyl	92	ND	57§	
16	Nevirapine	(standard)	1	0.8	0.0335/0.10¶	

Only one compound (10, Table 1) out of the approximately 65 first generation analogues prepared showed significant activity both in the enzymatic and the whole cell assay. Table 1 shows that the specific combination of 2-chlorobenzaldehyde and cyclohexyl isocyanide is required for potency, as other isocyanide combinations with this aldehyde gave poor inhibition results (cf. 10 with 2 and 6). Compound 10 acted as the lead compound for preparation of second-generation analogues, in which the isocyanide was kept constant and variously halogenated benzaldehydes were tested (Table 2).

From the enzyme inhibition results (Table 2), there is a clear preference for the 2-halo substituent in all cases of monosubstituted systems (10, 17 - 23). The size of the halogen has a strong influence when placed at position 4 (21 - 23).

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Table 2. Selected second generation analogues ( $R_1 = H$ ,  $R_3 = cyclohexyl$ )

No.	R <sub>2</sub>	% Residual Enzyme Activity	IC <sub>50</sub> (μM) MAGI Assay	No.	R <sub>2</sub>	% Residual Enzyme Activity
17	2-Fluorophenyl	14	18	23	4-Bromophenyl	100
18	2-Bromophenyl	30	2	24	2,6-Difluorophenyl	18
19	3-Chlorophenyl	76	ND	25	2,6-Dichlorophenyl	56
20	3-Bromophenyl	84	ND	26	2,4-Dichlorophenyl	40
21	4-Fluorophenyl	45	ND	27	3-Chloro-2-fluorophenyl	50
22	4-Chlorophenyl	78	ND	28	2,4,5-Trifluorophenyl	19

The addition of a second halogen to the 2-halo system at the 3-, 4- or 6-position is deleterious for any halogen except fluorine (cf. 24 and 28 with 25 - 27). The whole-cell results show unexpectedly good results for 2bromophenyl (18), but there is still clear preference for chlorine (10) over bromine. Hence, refinement using the preferred 2-chlorophenyl group at R2 was undertaken, varying the cycloalkyl group at R3 and introducing substituents on the 2-aminopyridine-derived component.

Table 3. Lead optimization

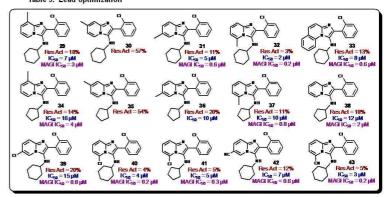
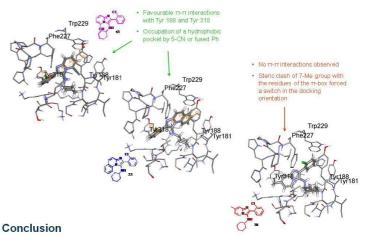


Table 3 shows that substitution at position 5 or 6 is highly favoured (31, 32, 36, 37, 39, 40-43), while that at 8 is tolerated (29 and 34). However, substitution at position 7 results in a dramatic loss in potency (30 and 35). Interestingly, the imidazo[1,2-a]quinoline (33) with ring fusion at positions 5 and 6 shows good activity, while the isoquinoline analogue with ring fusion at position 7 and 8 is completely inactive. A possible molecular explanation for this substitution preference is illustrated below. Cyclopentyl and cyclohexyl derivatives gave very similar results. Compounds substituted with 5-chloro, -cvano or -methyl groups showed whole-cell anti-HIV activity comparable to that of nevirapine (16, Table 1).



A large number of imidazo[1,2-a]pyridines were synthesised using the Groebke reaction and tested for anti-HIV activity. Compounds prepared from 2-chlorobenzaldehyde, cycloalkyl isocyanide and 6substituted 2-aminopyridines showed excellent inhibitory activity.

The percentage RT activity remaining after addition of the inhibitor PBMC (peripheral blood mononuclear cell) assay uses human PBMCs infected with HIV to test compounds for the ability to inhibit