

Available online at www.sciencedirect.com



Journal of Ethnopharmacology 92 (2004) 177-191



www.elsevier.com/locate/jethpharm

In vitro antiplasmodial activity of medicinal plants native to or naturalised in South Africa

Cailean Clarkson^a, Vinesh J. Maharaj^b, Neil R. Crouch^{c,d}, Olwen M. Grace^c, Pamisha Pillay^b, Motlalepula G. Matsabisa^e, Niresh Bhagwandin^e, Peter J. Smith^{a,*}, Peter I. Folb^a

^a Pharmacology Division, Department of Medicine, University of Cape Town, K-45 OMB GSH, Observatory 7925, South Africa

^b Bio/Chemtek, CSIR, P.O. Box 395, Pretoria 0002, South Africa

^c Ethnobotany Unit, National Botanical Institute, P.O. Box 52099, Berea Road, Durban 4007, South Africa

^d Natural Products Research Group, School of Chemistry, University of KwaZulu-Natal, Durban 4041, South Africa

^e Medical Research Council, P.O. Box 19070, Tygerberg 7505, South Africa

Received 13 November 2003; received in revised form 9 February 2004; accepted 9 February 2004

Available online 20 April 2004

Abstract

The increasing prevalence and distribution of malaria has been attributed to a number of factors, one of them being the emergence and spread of drug resistant parasites. Efforts are now being directed towards the discovery and development of new chemically diverse antimalarial agents. The present study reports on the in vitro antiplasmodial activity of 134 plant taxa native to or naturalised in South Africa, representing 54 families, which were selected semi-quantitatively using weighted criteria. The plant extracts were tested for in vitro activity against a *Plasmodium falciparum* strain D10 using the parasite lactate dehydrogenase (pLDH) assay. Of the 134 species assayed, 49% showed promising antiplasmodial activity (IC₅₀ \leq 10 µg/ml), while 17% were found to be highly active (IC₅₀ \leq 5 µg/ml). Several plant species and genera were shown for the first time to possess in vitro antiplasmodial activity. These results support a rational rather than random approach to the selection of antiplasmodial screening candidates, and identify a number of promising taxa for further investigation as plant-based antimalarial agents. © 2004 Elsevier Ireland Ltd. All rights reserved.

Keywords: Malaria; Antiplasmodial; Plasmodium falciparum; Ethnomedicinal plants; South Africa

1. Introduction

Despite intensive efforts to control malaria, the disease continues to be one of the greatest health problems facing Africa. It is estimated that there are at least 300 million clinical cases of malaria per annum, making it one of the top three killers among communicable diseases (WHO, 2003). Although a number of advances have been made towards the understanding of the disease, relatively few antimalarial drugs have been developed in the last 30 years (Ridley, 2002). Since the treatment and control of malaria depends largely on a limited number of chemoprophylactic and chemotherapeutic agents, there is an urgent need to develop novel, affordable antimalarial treatments. This urgency has been further highlighted by the increasing prevalence of drug resistant strains of the malaria parasite Plasmodium falciparum, which have contributed to the escalating disease burden.

Historically, the majority of antimalarial drugs have been derived from medicinal plants or from structures modelled on plant lead compounds. These include the quinoline-based antimalarials as well as artemisinin and its derivatives. Medicinal plants are commonly used in South African traditional healthcare to treat a range of ailments, including malaria and its associated symptoms (Watt and Breyer-Brandwijk, 1962). Of the 24,300 higher plant taxa recorded in the Flora of Southern Africa (FSA) region, approximately 15% are used for medicinal purposes (Arnold et al., 2002) and have been documented in various publications and electronic databases. The importance of the region's diverse medicinal plants lies not only in their chemotherapeutic value in traditional healthcare but also in their potential as sources of new chemical entities for drug discovery.

South Africa boasts remarkable biodiversity and rich cultural traditions of plant use. Scientific understanding of medicinal plants is, however, largely unexplored and pharmacological investigation of the South African flora only gained momentum recently (Van Wyk, 2002).

^{*} Corresponding author. Tel.: +27-21-4066289; fax: +27-21-4481989. *E-mail address:* psmith@uctgsh1.uct.ac.za (P.J. Smith).

In light of this and the pressing need for new antimalarial agents, the South African Department of Arts, Culture, Science and Technology (now the Department of Science and Technology) awarded an innovation fund to five South African institutions to evaluate local medicinal plants for antimalarial activity. The aim of this collaborative project was to discover novel, effective plant-based medicines for the treatment of malaria. In this article, we report on the in vitro antiplasmodial activity of 134 species of plants and identify potential sources of new antimalarial drugs.

2. Materials and methods

2.1. Selection and collection of plant material

A survey of relevant literature (30 books) on medicinal plant use in East and Southern Africa revealed approximately 700 taxa associated with malaria and/or fever. All taxa (623) occurring, indigenous or naturalised within the FSA region, were flagged as potential candidates for a targeted antiplasmodial screen. They were subsequently ranked following the application of weighted criteria, principally ethnobotanical and chemotaxonomic. Similar methodology has previously been applied to the selection of plant molluscicidal candidates from the FSA region (Clark et al., 1997). In the present study, this semi-quantitative approach objectively optimised the selection process and arguably maximised the likelihood of identifying positive antiplasmodial leads. Scores were based on each taxon's association with malaria and/or fever, the documented chemotherapeutic (antiplasmodial) potential of the plant family, use in traditional medicine, occurrence in the regional malaria-endemic area, and popularity in the local ethnomedicinal plant trade. Extra weighting was provided to plants indigenous to the FSA region. From the ranked list, 475 taxa attained total scores of seven or more, of which 134 species were collected throughout South Africa and subsequently investigated. Notably, the top-ranked taxon possessed a total score of 17 out of a possible maximum 20 points. The relevance and suitability of the selection approach and the scoring system will be reported on in due course in a separate publication. Voucher specimens were identified and deposited at the National Herbarium (Pretoria).

2.2. Preparation of extracts

Plant samples were separated into different components and dried in an oven at 30–60 °C. The drying time and temperature varied depending on the nature of the plant part. Dried plant material was ground to a coarse powder using a hammer mill and stored at ambient temperature prior to extraction. For each extraction procedure, 100–500 g of powdered plant material was sequentially extracted, typically with cold dichloromethane (DCM), DCM/methanol (MeOH) (1:1), MeOH and purified water. Organic extracts were concentrated by rotary vacuum evaporation below 45 °C and then further dried in vacuo at ambient temperature for 24 h. The aqueous extracts were concentrated by freeze-drying. All dried extracts were stored at -20 °C and the yields of the extracts, in terms of starting plant material, were recorded.

2.3. In vitro antiplasmodial activity

A chloroquine-sensitive strain (D10) of Plasmodium falciparum was continuously cultured according to the methods described by Trager and Jensen (1976), and parasite lactate dehydrogenase (pLDH) activity was used to measure parasite viability (Makler et al., 1995). The in vitro assays were performed as previously described by Clarkson et al. (2003). The IC_{50} values were obtained from the dose-response curves, using non-linear dose-response curve fitting analyses with GraphPad Prism v.3.00 software. Chloroquine diphosphate (Sigma) served as the positive control and was made up in Millipore water and serially diluted in medium to the required concentrations. All crude plant extracts were stored at -20 °C prior to testing and stock solutions were made up a day before the experiment and stored at -20 °C. Crude plant extracts were first dissolved in MeOH or dimethylsulphoxide (DMSO), depending on their solubility, sonicated for 10 min and then diluted in Millipore water to prepare a 2 mg/ml solution. The 2 mg/ml solution was further diluted in medium to give 200 µg/ml stock solutions. The highest concentration of solvent that the parasites were exposed to was 0.5%, which was shown to have no measurable effect on parasite viability. Extracts were tested in nine serial twofold dilutions (final concentration range: 100-0.2 µg/ml) in 96-well microtitre plates. All tests were performed in duplicate and no attempt was made to determine 50% inhibitory concentration (IC₅₀) values in excess of 100 µg/ml.

3. Results and discussion

For the purpose of this study, an IC₅₀ value of $\leq 10 \,\mu$ g/ml was classified as promising activity, and $\leq 5 \,\mu$ g/ml was considered to be highly active. These two concentrations were chosen with the reasoning that inhibition of parasite growth at the low concentrations would indicate selective activity as opposed to higher concentrations where non-specific toxicity is often observed. A total of 134 plant taxa, representing 54 families, were tested for in vitro antiplasmodial activity (Table 1). Sixty-six species showed promising antiplasmodial activity with IC₅₀ values of $\leq 10 \,\mu$ g/ml, of which 23 were found to be highly active with IC₅₀ values of $\leq 5 \,\mu$ g/ml (Table 2). Due to the large volumes of data generated in this study, only the highly active extracts are highlighted (Table 2) and further discussed.

Although a substantial amount of phytochemical research has been carried out on *Catha edulis* (Vahl) Forssk. ex Endl.

Table 1	
In vitro antiplasmodial activity of the plant extracts against Plasmodium falciparum D	10

Family	Plant species	Voucher number	Plant part	Solvent	% Yield	IC ₅₀ (µg/ml)
Acanthaceae	Asystasia gangetica T. Anderson	BP00901	Twigs	DCM/MeOH (1:1) Water	0.3 7.6	16 > 100
		BP00901	Leaves	DCM/MeOH (1:1) Water	0.9 1.1	7 > 100
	Justicia flava (Vahl) Vahl	BP00960	Whole plant	DCM/MeOH (1:1) Water	0.6 3.7	31 > 100
Amaranthaceae	Achyranthes aspera L.	BP01260	Whole plant	DCM/MeOH (1:1) Water	0.6 0.8	9.9 > 100
Amaryllidaceae	Crinum macowanii Baker	BP00907	Bulbs	DCM/MeOH (1:1)	0.7	26
				Water	1.1	25
Anacardiaceae	Lannea discolor (Sond.) Engl.	BP01389	Fruit	DCM MeOH/DCM Water	0.4 0.6 1.1	25 > 100 > 100
Annonaceae	Annona senegalensis Pers. subsp. senegalensis	EN00334	Leaves	DCM	0.7	35
	Artabotrys brachypetalus Benth.	BP00210	Leaves/twigs	DCM/MeOH (1:1) DCM/MeOH (1:1) Water	0.9 0.9	45 > 100 > 100
	Artabotrys monteiroae Oliv.	BP01273	Twigs	Water DCM/MeOH (1:1) Water	1.3 1.2 4.8	> 100 8.7 > 100
		BP01273	Leaves	DCM/MeOH (1:1) Water	1.3 0.7	22 23
Apiaceae	Alepidea amatymbica Eckl. & Zeyh.	EN00994	Whole plant	DCM/MeOH (1:1)	0.7	12.5
	Berula erecta (Huds.) Coville	BP00957	Whole plant	Water DCM/MeOH (1:1)	1.7 0.3	> 100 6.6
	Centella asiatica (L.) Urb.	BP00955	Leaves	Water DCM/MeOH (1:1)	2.7 0.4	> 100 8.3
Apocynaceae	Carissa edulis Vahl	EN00097	Stems	DCM	0.4	33
. ipoegnaeeae		2111000077	Stellis	DCM/MeOH (1:1)	0.9	60
	Carissa edulis Vahl	EN00097	Stems	MeOH Water	0.5 1.5	100 > 100
		EN00097	Seeds	DCM	0.7	> 100 > 100
				DCM/MeOH (1:1)	0.9	> 100
				MeOH	0.9	> 100
		-		Water	0.6	> 100
		EN00097	Leaves	DCM	1.1	100
				DCM/MeOH (1:1) MeOH	0.8 0.9	> 100 > 100
				Water	1.3	> 100 > 100
	Diplorhynchus condylocarpon (Müll. Arg.) Pichon	EN00300	Roots	DCM	0.6	> 100
				DCM/MeOH (1:1)	0.9	24
				Water	0.8	> 100
	Rauvolfia caffra Sond.	EN00232	Fruit	DCM	0.8	26.5
				DCM/MeOH (1:1)	0.3	55
		EN100222	Deete	Water	1.1	> 100
		EN00232	Roots	DCM DCM/MeOH (1:1)	1.3 0.9	88 67
				Water	1.2	90
	Gomphocarpus fruticosus (L.) Aiton. f.	BP01151	Fruit	DCM/MeOH (1:1)	0.6	90 26
Araliaceae	Cussonia spicata Thunb.	BP00031	Leaves	DCM	1.6	45
				DCM/MeOH (1:1)	0.9	13
				MeOH	1.0	27.5
				Water	1.8	90
		EN00867	Fruit	DCM/MeOH (1:1)	0.4	14
				Water	1.2	> 100

Family	Plant species	Voucher number	Plant part	Solvent	% Yield	IC ₅₀ (µg/ml)
	Schefflera umbellifera (Sond.) Baill.	EN00060	Leaves	DCM	0.6	3.7
				DCM/MeOH (1:1) MeOH	0.6 0.9	19.5 49.5
	<i>Schefflera umbellifera</i> (Sond.) Baill.	EN00060	Leaves	Water	1.2	> 100
	Daili.	EN00060	Roots	DCM	1.1	7.5
				DCM/MeOH (1:1)	0.9	5.8
				MeOH	0.7	> 100
				Water	1.0	> 100
		EN00060	Stems	DCM	0.8	42
				DCM/MeOH (1:1)	0.9	15
				MeOH	0.6	35
				Water	0.9	100
				Water	1.2	> 100
		BP01151	Leaves/twigs	DCM/MeOH (1:1)	1.2	25
			-	Water	0.9	> 100
	<i>Xysmalobium undulatum</i> (L.) Aiton. f.	BP01179	Whole plant	DCM/MeOH (1:1)	1.4	6
				Water	1.8	> 100
sparagaceae	Asparagus virgatus Baker	BP01029	Whole plant	DCM/MeOH (1:1)	0.7	8
				Water	0.2	> 100
Asphodelaceae	Aloe ferox Mill.	BP00469	Whole plant	DCM/MeOH (1:1)	0.7	8
				Water	1.1	> 100
		EN00588	Fruit	DCM	0.4	20
				DCM/MeOH (1:1)	0.8	14
				Water	0.6	18
		EN00588	Stems	DCM	0.8	30
				DCM/MeOH (1:1)	0.8	15.5
				Water	0.9	> 100
		EN00588	Roots	DCM	0.7	13
				DCM/MeOH (1:1)	0.8	8.5
				Water	0.5	> 100
	Aloe ferox Mill.	EN00588	Leaves	DCM	0.8	21
				DCM/MeOH (1:1)	0.4	> 100
				Water	0.7	> 100
	Aloe maculata All.	BP01314	Whole plant	DCM/MeOH (1:1)	0.6	12.4
				Water	3.0	> 100
	Aloe marlothii A. Berger	BP00049	Leaves	DCM	0.8	74
	-			DCM/MeOH (1:1)	0.6	90
				MeOH	0.6	> 100
				Water	0.8	71
		BP00049	Whole plant	DCM	0.4	3.5
			•	DCM/MeOH (1:1)	0.7	18
				Water	1.0	> 100
	Ageratum conyzoides L.	BP01233	Whole plant	DCM/MeOH (1:1)	1.4	27
	0		Ĩ	Water	0.9	> 100
steraceae	Artemisia afra Jacq. ex Willd.	EN00148	Leaves	DCM	0.7	5
				DCM/MeOH (1:1)	0.9	7.3
				MeOH	0.9	8
				Water	0.6	> 100
	Bidens pilosa L.	EN00001	Leaves	DCM	0.4	8.5
				DCM/MeOH (1:1)	0.8	11
				MeOH	0.6	5
				Water	1.6	70
	Conyza albida Spreng.	BP01221	Whole plant	DCM/MeOH (1:1)	0.7	2
				Water	1.1	> 100
	Conyza podocephala DC.	BP00378	Whole plant	DCM/MeOH (1:1)	0.6	6.8
				Water	1.3	> 100
	Conyza scabrida DC.	BP00443	Flowers	DCM/MeOH (1:1)	0.3	7.8
				Water	0.4	> 100

Table 1 (Co	ontinued)
-------------	-----------

amily	Plant species	Voucher number	Plant part	Solvent	% Yield	IC ₅₀ (µg/ml)
	Conyza scabrida DC.	BP00443	Leaves	DCM/MeOH (1:1) Water	0.6 2.2	11.5 > 100
		BP00443	Twigs	DCM/MeOH (1:1) Water	0.7 1.1	> 100 11 > 100
	Helichrysum nudifolium (L.) Less.	EN01023	Whole plant	DCM/MeOH (1:1)	0.5	> 100 6.8
	Osteospermum imbricatum L.	EN00874	Stems	Water DCM/MeOH (1:1)	2.4 1.2	> 100 7.3
	Pentzia globosa Less.	EN00506	Leaves	Water DCM	0.7 0.7	> 100 12.5
		EN00506	Stome	DCM/MeOH (1:1) Water DCM	0.6 1.2 0.9	19.5 53 9.5
		EN00506	Stems	DCM DCM/MeOH (1:1) Water	0.9 0.9 0.6	9.5 15.5 > 100
		EN00393	Roots	DCM DCM/MeOH (1:1) Water	1.1 0.8 0.8	> 100 8 14 > 100
	<i>Psiadia punctulata</i> (DC.) Oliv. & Hiern ex Vatke	CS00005	Twigs	DCM	0.3	9
		CN00044	Leaves	Water DCM DCM/MeOH (1:1) Water	0.9 1.2 1.4	> 100 14 22.5
		BP00278	Whole plant	water DCM/MeOH (1:1) Water	0.6 1.2 0.9	> 100 18 > 100
	Senecio oxyriifolius DC.	BP01037	Whole plant	DCM/MeOH (1:1) Water	0.7 2.6	13 > 100
	Spilanthes mauritiana (Pers.) DC.	EN00075	Stems	DCM	0.8	38
	Spilanthes mauritiana (Pers.)	EN00075	Stems	DCM/MeOH (1:1) MeOH	0.9 0.8	5.3 64
	DC.	EN00075	Stellis	Water	2.6	> 100
	Tarchonanthus camphoratus L.	BP00263	Whole plant	DCM/MeOH (1:1) Water	0.6 0.9	> 100 6 > 100
		EN00389	Leaves	DCM DCM/MeOH (1:1) Water	0.9 1.2 1.8	> 100 7.5 13 > 100
		EN00389	Roots	DCM DCM/MeOH (1:1)	1.8 1.1 0.8 0.7	> 100 60 24 > 100
	Tridax procumbens L.	BP00892	Whole plant	Water DCM/MeOH (1:1) Water	0.7 0.4 1.9	> 100 17 > 100
	<i>Vernonia colorata</i> (Willd.) Drake subsp. <i>colorata</i>	BP01279	Twigs	DCM/MeOH (1:1)	0.4	14.1
		BP01279	Leaves	Water DCM/MeOH (1:1) Water	0.6 1.5 1.0	> 100 4.7 87.7
	Vernonia fastigiata Oliv. & Hiern	EN00118	Leaves	DCM DCM/MeOH (1:1) MeOH Water	1.0 0.8 0.6 0.9	10 15 30 > 100
	<i>Vernonia hirsuta</i> (DC.) Sch. Bip. ex Walp.	EN01012	Whole plant	DCM/MeOH (1:1)	0.7	14
	Vernonia myriantha Hook. f.	EN00044	Roots	Water DCM DCM/MeOH (1:1) MeOH	1.2 0.9 0.8 0.8	> 100 61 37.5 28
		EN00044	Leaves	Water DCM DCM/MeOH (1:1)	2.3 0.6 0.8	70 3 13.5

Family	Plant species	Voucher number	Plant part	Solvent	% Yield	IC ₅₀ (µg/ml)
	Vernonia myriantha Hook. f. Vernonia natalensis Sch. Bip. ex Walp.	EN00044 EN00331	Leaves Whole plant	Water DCM	2.1 1.5	> 100 19.5
	L			DCM/MeOH (1:1) Water	0.7 1.1	24 > 100
	Vernonia oligocephala (DC.)	EN00314	Roots	DCM	0.7	> 100
	Sch. Bip. ex Walp.			DCM/MeOH (1:1)	0.7	20
				Water	2.1	> 100
		EN00314	Leaves	DCM	1.2	3.5
				DCM/MeOH (1:1)	0.8	5.5
				Water	2.2	> 100
Bignoniaceae	Kigelia africana (Lam.) Benth.	EN00217	Leaves	DCM	0.7	51
				DCM/MeOH (1:1)	0.8	87
		5501000		Water	1.7	> 100
	<i>Tecomaria capensis</i> (Thunb.) Lindl.	BP01302	Leaves	DCM/MeOH (1:1)	0.1	11.6
		DD 01202	Thereit and	Water	3.1	10.9
		BP01302	Twigs	DCM/MeOH (1:1) Water	0.1 2.0	81.9 10.2
Capparaceae	Capparis tomentosa Lam.	EN00222	Leaves	DCM	0.5	65
Capparaceae	Cuppans iomeniosa Lani.	LINOUZZZ	Leaves	DCM/MeOH (1:1)	0.9	> 100
				Water	0.7	> 100
		EN00222	Stems	DCM	0.4	41.5
		LITTOOLLE	Stellis	DCM/MeOH (1:1)	0.7	> 100
				Water	0.9	> 100
		EN00222	Roots	DCM	0.4	38
		21100222	1000	DCM/MeOH (1:1)	0.8	> 100
				Water	1.2	> 100
Celastraceae	<i>Catha edulis</i> (Vahl) Forssk. ex Endl.	EN00159	Seeds	DCM	0.3	46
	<i>Catha edulis</i> (Vahl) Forssk. ex Endl.	EN00159	Seeds	DCM/MeOH (1:1)	0.4	10
				MeOH	0.4	78.5
				Water	0.3	> 100
		EN00159	Roots	DCM	0.6	0.63
				DCM/MeOH (1:1)	0.8	4.8
				MeOH	0.6	23.5
		F3404.50		Water	1.2	> 100
		EN00159	Leaves	DCM	0.9	0.77
				DCM/MeOH (1:1)	1.1	6.9
				MeOH	1.0	7.7
	Maytenus senegalensis (Lam.)	EN00218	Roots	Water DCM	1.4 2.3	> 100 15.5
	Exell.			Water	1.2	> 100
		EN00218	Stems	DCM	2.7	> 100 42
		EIN00218	Stellis	DCM/MeOH (1:1)	1.3	48.3
				Water	0.4	> 100
	<i>Maytenus undata</i> (Thunb.) Blakelock	EN00029	Leaves	DCM	0.5	> 100
	Burelock			DCM/MeOH (1:1)	0.7	21
				MeOH	0.7	60
				Water	1.1	> 100
		EN00029	Stems	DCM	0.5	85
				DCM/MeOH (1:1)	0.6	24
				MeOH	0.5	38
				Water	2.1	> 100
		EN00029	Roots	DCM	0.6	23
				DCM/MeOH (1:1)	0.8	36
				MeOH	0.6	40
				Water	2.1	> 100

Family	Plant species	Voucher number	Plant part	Solvent	% Yield	IC ₅₀ (μg/ml)
Chrysobalanaceae	<i>Parinari curatellifolia</i> Planch. ex Benth.	EN00065	Leaves/flowers	DCM	0.8	17
				DCM/MeOH (1:1)	0.7	40
				MeOH	0.7	46.5
				Water	1.6	81
		EN00065	Roots	DCM	0.6	5.3
				DCM/MeOH (1:1)	1.2	22.5
				MeOH	0.9	30.5
				Water	1.9	63.5
Clusiaceae	Hypericum aethiopicum Thunb.	BP00365	Leaves/flowers	DCM/MeOH (1:1)	0.7	1.4
Clustaceae	Hypericum deiniopicum Thuno.	BI 00505	Leaves/nowers	Water	1.4	90
Colchicaceae	Gloriosa superba L.	BP01040	Whole plant	DCM/MeOH (1:1)	0.8	17
Colemeaceae	Gioriosa superba E.	DI 01040	whole plan	Water	3.7	> 100
Combretaceae	Combretum zeyheri Sond.	BP01239	Twigs	DCM/MeOH (1:1)	0.4	15
compretaceae	Comprehant Leynert Bond.	BIOILSY	1.1185	Water	1.2	> 100
Cucurbitaceae	Momordica balsamina L.	FP00062	Whole plant	DCM/MeOH (1:1)	0.9	18
			T	Water	2.1	> 100
		MM00075	Stem	DCM/MeOH (1:1)	0.7	5.3
				Water	0.8	> 100
		MM00075	Leaves	DCM/MeOH (1:1)	0.7	6
		DD01055		Water	2.3	> 100
	Zehneria scabra (L.f.) Sond. subsp. scabra	BP01065	Whole plant	DCM/MeOH (1:1)	0.9	5.6
	I I I I I I I I I I I I I I I I I I I			Water	3.8	> 100
Ebenaceae	Euclea natalensis A.DC.	EN00760	Stems	DCM/MeOH (1:1)	1.4	5.3
				Water	2.7	> 100
		EN00760	Roots	DCM/MeOH (1:1)	0.9	5.1
				Water	1.3	> 100
	Euclea undulata Thunb.	BP01105	Leaves	DCM/MeOH (1:1)	1.7	11
				Water	1.1	> 100
	Euclea undulata Thunb.	BP01105	Twigs	DCM/MeOH (1:1)	1.2	4.6
				Water	0.4	> 100
Euphorbiaceae	<i>Bridelia micrantha</i> (Hochst.) Baill.	BP01316	Twigs	DCM/MeOH (1:1)	0.5	59.3
				Water	1.1	6.9
	Clutia hirsuta E. Mey. ex Sond.	BP01193	Whole plant	DCM/MeOH (1:1)	1.0	15
				Water	2.3	50
	<i>Croton gratissimus</i> Burch. var. <i>subgratissimus</i> (Prain) Burtt Davy	BP00929	Leaves	DCM	0.7	3.5
				DCM/MeOH (1:1)	1.2	11.5
				МеОН	1.1	29
				Water	2.3	95
	Croton menyhartii Pax	BP01213	Leaves	DCM/MeOH (1:1)	2.0	1.7
		=		Water	2.0	> 100
		BP01213	Twigs	DCM/MeOH (1:1)	0.8	15
			J	Water	0.7	> 100
	Euphorbia heterophylla L.	BP01249	Whole plant	DCM/MeOH (1:1)	1.3	40
			-	Water	4.8	> 100
	Euphorbia tirucalli L.	RP00008	Leaves	DCM	0.8	12
				DCM/MeOH (1:1)	1.2	23.5
				MeOH	0.7	> 100
				Water	2.3	83
	<i>Flueggea virosa</i> (Roxb. ex Willd.)	BP00207	Leaves/twigs	DCM/MeOH (1:1)	0.3	19
	Voigt subsp. <i>virosa</i>			Water	1.4	11.4
	Ricinus communis L. var.	EN00768	Leaves	DCM/MeOH (1:1)	1.4	27.5
	communis					
	communis				· ·	
	community	EN00768	Stems	Water DCM/MeOH (1:1)	0.6 0.9	> 100 8

Family	Plant species	Voucher number	Plant part	Solvent	% Yield	IC ₅₀ (µg/ml)
				Water	1.3	> 100
		EN00768	Fruit	DCM/MeOH (1:1)	0.5	90
				Water	0.6	> 100
abaceae	Acacia nilotica (L.) Willd. ex Delile	BP01209	Twigs	DCM/MeOH (1:1)	0.4	13
	Acacia nilotica (L.) Willd. ex Delile	BP01209	Twigs	Water	6.8	32
	Acacia tortilis (Forssk) Hayne	BP01203	Whole plant	DCM/MeOH (1:1) Water	0.7 0.1	4.8 > 100
	Crotalaria burkeana Benth.	EN00126	Leaves	DCM	0.7	30
	Crotataria barkeana Bonti.	21100120	Leaves	DCM/MeOH (1:1)	1.3	50
				МеОН	1.5	> 100
				Water	0.8	> 100
		EN00126	Roots	DCM	0.5	9.5
		21100120	Roots	DCM/MeOH (1:1)	0.9	13
				МеОН	1.3	> 100
				Water	1.1	> 100
	Elephantorrhiza elephantina	BP01010	Roots	DCM/MeOH (1:1)	0.4	28
	(Burch.) Skeels					
				Water	1.5	> 100
		BP01010	Leaves	DCM/MeOH (1:1)	0.1	26
				Water	1.2	> 100
	Parkinsonia aculeata L.	BP01144	Twigs	DCM/MeOH (1:1)	0.3	9
				Water	0.8	> 100
	Piliostigma thonningii (Schumach.) Milne-Redh.	BP01294	Leaves	DCM/MeOH (1:1)	0.9	32
				Water	1.0	> 100
		BP01294	Fruit	DCM/MeOH (1:1)	0.3	32.4
				Water	4.1	59
		BP01294	Twigs	DCM/MeOH (1:1)	0.3	25.9
				Water	0.8	> 100
	<i>Pseudarthria hookeri</i> Wight & Arn. var. <i>hookeri</i>	BP01217	Leaves	DCM/MeOH (1:1)	1.3	100
				Water	4.8	> 100
	Pterocarpus angolensis DC.	EN00083	Stems	DCM	0.7	15
				DCM/MeOH (1:1)	1.1	60
				MeOH	0.9	71
	Pterocarpus angolensis DC.	EN00083	Stems	Water	1.8	> 100
	1 0	EN00083	Roots	DCM	0.9	10.6
				DCM/MeOH (1:1)	0.9	25.5
				МеОН	1.5	33.5
				Water	2.0	80
	Senna didymobotrya (Fresen.) Irwin & Barneby	BP01078	Leaves	DCM/MeOH (1:1)	1.7	40
				Water	3.8	> 100
		BP01078	Twigs	DCM/MeOH (1:1)	0.7	9.5
			0	Water	1.6	> 100
		BP01078	Pods	DCM/MeOH (1:1)	0.3	18
				Water	2.5	> 100
	Senna petersiana (Bolle) Lock	BP01290	Leaves	DCM/MeOH (1:1)	2.7	> 100
	/			Water	6.2	> 100
		BP01290	Twigs	DCM/MeOH (1:1)	0.3	13
			-	Water	0.7	> 100
acourtiaceae	Flacourtia indica (Burm. f.)	EN0029	Roots	DCM	0.7	86.5
	Merr.			DCM/MeOH (1:1)	1.3	78
				Water	0.9	> 100
antionasas-	Anthoplaista anon dia an Oil-	NDOOOO	Lacres			
entianaceae	Anthocleista grandiflora Gilg	NB00028	Leaves	DCM	1.4	> 100
				DCM/MeOH (1:1)	1.2	> 100
	Delanomium delaniti i a	DD 01010	W/k -1 1	Water	2.6	90 15
	Pelargonium alchemilloides (L.)	BP01018	Whole plant	DCM/MeOH (1:1)	0.7	15

Family	Plant species	Voucher number	Plant part	Solvent	% Yield	IC ₅₀ (µg/ml)
				Water	5.5	100
Goodeniaceae	Scaevola plumieri (L.) Vahl	BP01075	Twigs	DCM	0.9	11
				Water	15.3	> 100
Iyacinthaceae	<i>Eucomis autumnalis</i> (Mill.) Chitt.	NB00025	Bulbs	DCM	1.8	70
				DCM/MeOH (1:1)	2.2	9.5
				Water	3.6	> 100
llecebraceae	Pollichia campestris Aiton	BP00559	Twigs	DCM/MeOH (1:1)	0.5	6.8
			0	Water	0.7	> 100
		BP00416	Whole plant	DCM/MeOH (1:1)	2.3	25
				Water	3.2	> 100
		EN00124	Leaves	DCM	1.7	23
				DCM/MeOH (1:1)	1.6	17
				MeOH	0.9	70
				Water	2.2	> 100
		EN00124	Fruit	DCM	0.3	14
				DCM/MeOH (1:1)	0.3	27
				MeOH	0.5	68
				Water	0.4	> 100
Kirkiaceae	Kirkia wilmsii Engl.	BP01242	Leaves	DCM/MeOH (1:1)	1.3	3.7
	Ũ			Water	2.5	> 100
r	$H_{\rm eff}$ is a set in set of $(\mathbf{L}_{\rm eff})$ $\mathbf{D}_{\rm eff}$	DD00242	T / . t / f t	$DCM/M_{\odot}OU(1,1)$	0.9	17.5
Lamiaceae	Hyptis pectinata (L.) Poit.	BP00243	Leaves/stems/fruit	DCM/MeOH (1:1)	0.8	17.5
	Leonotis leonurus (L.) R.Br.	EN00809	Roots	Water DCM/MeOH (1:1)	0.7 0.8	> 100 15
	Leonous teonurus (L.) K.BI.	EN00809	ROOIS	Water	0.8 1.1	> 100
		BP00444	Twigs	DCM/MeOH (1:1)	0.9	> 100
		DI 00444	1 wigs	Water	1.0	> 100
		BP00444	Leaves	DCM/MeOH (1:1)	0.8	5.4
		DI 00444	Leaves	Water	1.3	> 100
	Leonotis nepetifolia (L.) R.Br.	BP01247	Whole plant	DCM/MeOH (1:1)	0.6	15
	(_)		······	Water	0.9	> 100
	Leonotis ocymifolia (Burm. f.) Iwarsson	BP01257	Leaves	DCM	1.3	17
				DCM/MeOH (1:1)	0.9	12
				MeOH	0.6	6.1
				Water	0.8	> 100
	<i>Leonotis ocymifolia</i> (Burm. f.) Iwarsson	BP01257	Fruit	DCM	0.3	38
				DCM/MeOH (1:1)	0.6	20
				MeOH	0.3	40
				Water	0.4	> 100
		BP01257	Roots	DCM	0.8	17
				DCM/MeOH (1:1)	0.9	28
				MeOH	1.2	> 100
	Laphotic acumitalia (Dum f)	EN01257	Whole plant	Water DCM/MeOH (1:1)	2.6 0.8	> 100 28
	<i>Leonotis ocymifolia</i> (Burm. f.) Iwarsson var. <i>raineriana</i> (Vis.) Iwarsson	EN01257	Whole plant		0.8	28
				Water	1.5	> 100
	Leucas martinicensis (L.) R.Br.	BP01204	Whole plant	DCM/MeOH (1:1)	0.8	13.3
			•	Water	0.7	> 100
	Ocimum americanum L. var. americanum	BP01210	Whole plant	DCM/MeOH (1:1)	0.1	4.2
				Water	0.8	> 100
	Salvia repens Burch. ex Benth.	BP00998	Whole plant	DCM/MeOH (1:1)	1.0	10.8
				Water	1.3	> 100
	<i>Tetradenia riparia</i> (Hochst.) Codd	EN00341	Leaves	DCM	0.9	> 100
				DCM/MeOH (1:1)	0.7	> 100
				Water	2.6	> 100

Family	Plant species	Voucher number	Plant part	Solvent	% Yield	IC ₅₀ (µg/ml)
Lecythidaceae	Barringtonia racemosa (L.) Roxb.	BP00919	Leaves	DCM/MeOH (1:1)	1.2	18
				Water	4.0	70
		BP00919	Twigs	DCM/MeOH (1:1)	0.5	5.7
			U	Water	1.7	> 100
Iaesaceae	Maesa lanceolata Forssk.	BP01226	Twigs	DCM/MeOH (1:1)	0.7	5.9
			-	Water	0.2	> 100
Ieliaceace	Ekebergia capensis Sparrm.	BP00442	Fruit	DCM/MeOH (1:1)	0.4	10
				Water	0.6	> 100
		BP00442	Twigs	DCM/MeOH (1:1)	0.5	18
	Ekebergia capensis Sparrm.	BP00979	Twigs	Water	0.8	> 100
	Trichilia emetica Vahl	BP00958	Leaves/twigs	DCM/MeOH (1:1)	3.4	3.5
	subsp. emetica					
		5501015	-	Water	2.2	> 100
	Turraea floribunda Hochst.	BP01265	Leaves	DCM/MeOH (1:1)	0.5	8.8
				Water	2.8	> 100
Iyrtaceae	Syzigium cordatum Hochst. ex Sond. var. cordatum	BP01268	Twigs	DCM/MeOH (1:1)	1.8	14.7
				Water	1.3	48.3
		BP01268	Leaves	DCM/MeOH (1:1)	0.7	22.8
				Water	0.9	23.6
Iyrsinaceae	Rapanea melanophloeos (L.)	BP01031	Leaves	DCM/MeOH (1:1)	2.0	44
	Mez			Water	1.2	> 100
		BP01031	Twigs	DCM/MeOH (1:1)	1.2	> 100 40
		BF01031	Twigs	Water	0.9	> 100
1	View in a floor Court and floor	EN00110	T			
lacaceae	Ximenia caffra Sond. var. caffra	EN00110	Leaves	DCM DCM/MeOH (1:1)	0.7 0.9	43.5 55
						100
				MeOH	0.7	
		EN100110	D (Water	2.3	> 100
		EN00110	Roots	DCM	0.5	> 100
				DCM/MeOH (1:1)	0.7	> 100
				MeOH Water	1.0 1.9	> 100 > 100
leaceae	<i>Olea europaea</i> L. subsp. <i>africana</i> (Mill.) P.S. Green	BP01236	Leaves	DCM/MeOH (1:1)	0.7	12
				Water	0.7	> 100
		BP01236	Twigs	DCM/MeOH (1:1)	1.2	13
			e	Water	1.1	> 100
ittosporaceae	Pittosporum viridiflorum Sims	EN00049	Whole plant	DCM	1.5	3
1			. I	DCM/MeOH (1:1)	0.9	10
				MeOH	0.6	27.7
	Pittosporum viridiflorum Sims	EN00049	Whole plant	Water	1.4	> 100
		EN00049	Leaves/flowers	DCM	0.8	28
		L1(0004)	Leaves/ nowers	DCM/MeOH (1:1)	0.6	20 47
				МеОН	0.8	70.5
				Water	1.7	> 100
antaginaceae	Plantago major L.	EN00499	Whole plant	DCM	0.9	21.5
anaginacouc	. tantago nagor L.	L1007))	more plunt	DCM/MeOH (1:1)	1.2	45
				Water	1.2	> 100
lumbaginaceae	Plumbago zeylanica L.	EN00208	Roots	DCM	0.8	43
	. amougo contante D.	11100200	10005	DCM/MeOH (1:1)	1.3	34
				MeOH (1.1)	1.5	54 77.3
				Water	2.4	> 100
		EN00208	Leaves	DCM	2.4 1.0	> 100
		EIN00208	Leaves			
				DCM/MeOH (1:1)	0.7 0.9	4.8 5.5
				MeOH Water		5.5 > 100
					1.9	

Family	Plant species	Voucher number	Plant part	Solvent	% Yield	IC ₅₀ (µg/ml)
Poaceae	<i>Cymbopogon validus</i> (Stapf) Stapf ex Burtt Davy	BP01227	Whole plant	MeOH/DCM (1:1)	0.9	5.8
				Water	0.8	> 100
	Setaria megaphylla (Steud.) T. Durand & Schinz	BP01200	Whole plant	MeOH/DCM (1:1)	0.8	4.5
				Water	0.9	> 100
Polygonaceae	Rumex crispus L.	EN00569	Leaves	DCM	1.2	36.8
				DCM/MeOH (1:1)	1.5	60
				Water	0.9	> 100
		EN00569	Roots	DCM	0.9	14
				DCM/MeOH (1:1)	0.7	32
		5501001		Water	1.8	> 100
	Rumex sagittatus Thunb.	BP01201	Whole plant	DCM/MeOH (1:1)	0.5	18
				Water	0.1	100
taeroxylaceae	<i>Ptaeroxylon obliquum</i> (Thunb.) Radlk.	EN00646	Roots	DCM	0.6	19
				DCM/MeOH (1:1)	0.9	17
				Water	1.3	> 100
		EN00646	Leaves	DCM	0.8	19.5
				DCM/MeOH (1:1)	0.5	22.8
				Water	0.6	> 100
		EN00646	Stems	DCM	0.7	11.5
				DCM/MeOH (1:1)	0.8	5.5
				Water	1.2	> 100
Ranunculaceae	Clematis brachiata Thunb.	BP00192	Leaves/stems/flowers	DCM/MeOH (1:1)	0.8	20
				Water	3.0	> 100
	Ranunculus multifidus Forssk.	BP00962	Whole plant	DCM/MeOH (1:1)	0.2	2.3
				Water	2.7	> 100
Rhamnaceae	Ziziphus mucronata Willd.	BP00005	Leaves	DCM	0.8	12
				DCM/MeOH (1:1)	1.3	> 100
				MeOH	0.4	> 100
				Water	0.8	> 100
Rhizophoraceae	Bruguiera gymnorhiza (L.) Lam.	BP01275	Twigs	DCM/MeOH (1:1)	0.6	11.7
				Water	0.9	> 100
		BP01275	Leaves	DCM/MeOH (1:1)	1.2	15.3
				Water	2.6	> 100
	Rhizophora mucronata Lam.	BP01274	Leaves	DCM/MeOH (1:1)	0.7	24
				Water	4.5	25
		BP01274	Twigs	DCM/MeOH (1:1)	0.6	5.6
				Water	2.2	> 100
Rubiaceae	Burchellia bubalina (L.f.) Sims	BP00936	Twigs	DCM/MeOH (1:1)	0.4	18
				Water	1.3	> 100
		BP00936	Leaves	DCM/MeOH (1:1)	0.8	50
	Cephalanthus natalensis Oliv.	BP01299	Leaves	DCM/MeOH (1:1)	0.2	24.3
				Water	3.6	> 100
		BP01299	Twigs	DCM/MeOH (1:1)	0.1	16.5
		DD00000		Water	1.4	> 100
	<i>Vangueria infausta</i> Burch. subsp. <i>infausta</i>	BP00004	Fruit	DCM/MeOH (1:1)	0.9	23
				Water	0.7	> 100
Rutaceae	Agathosma apiculata G. Mey.	BP01123	Whole plant	DCM/MeOH (1:1)	1.1	5.2
				Water	1.9	> 100
	<i>Agathosma puberula</i> (Steud.) Forc.	EN00697	Roots	DCM	0.8	33
				DCM/MeOH (1:1)	0.7	19
				Water	1.4	> 100
		EN00697	Stems	DCM	0.9	15
				DCM/MeOH (1:1)	0.8	15
				DCM/MCOII (1.1)	0.0	15

Family	Plant species	Voucher number	Plant part	Solvent	% Yield	IC ₅₀ (µg/ml)
	<i>Clausena anisata</i> (Willd.) Hook. f. ex Benth var. <i>anisata</i>	BP01125	Twigs	DCM/MeOH (1:1)	0.6	18
				Water	1.0	> 100
		BP01125	Leaves	DCM/MeOH (1:1)	1.0	55
				Water	2.9	> 100
	Diosma sp.	EN00735	Roots	DCM	0.4	10
				DCM/MeOH (1:1)	0.8	80
				Water	1.2	> 100
	<i>Macrostylis squarrosa</i> Bartl. & H.L. Wendl.	EN00758	Stems	DCM/MeOH (1:1)	0.6	10
				Water	0.7	> 100
Sapindaceae	Cardiospermum halicacabum L.	BP01286	Whole plant	DCM/MeOH (1:1)	1.5	20
				Water	2.8	> 100
	Dodonaea viscosa Jacq.	EN00791	Leaves	DCM/MeOH (1:1)	0.5	15.5
				Water	0.7	> 100
	<i>Hippobromus pauciflorus</i> (L.f.) Radlk.	BP01229	Leaves	DCM/MeOH (1:1)	0.4	34
	Hippobromus pauciflorus (L.f.) Radlk.	BP01229	Leaves	Water	0.2	60
		BP01229	Twigs	DCM/MeOH (1:1)	0.6	5.9
				Water	0.9	> 100
terculiaceae	Hermannia depressa N.E.Br.	BP01081	Whole plant	DCM/MeOH (1:1)	0.4	6.9
			Press	Water	4.1	70
	Waltheria indica L.	BP01241	Whole plant	DCM/MeOH (1:1)	0.7	> 100
	waimeria maica L.	DI 01241	whole plant	Water	2.2	> 100
trychnaceae	Strychnos madagascariensis Poir.	EN00286	Stems	DCM	0.8	70
				DCM/MeOH (1:1)	0.9	31.5
				Water	2.6	85
		EN00286	Leaves	DCM	0.5	65
				DCM/MeOH (1:1)	0.8	40
				Water	1.3	74
		EN00286	Roots	DCM	0.7	56
		21100200	10005	DCM/MeOH (1:1)	0.9	> 100
				Water	1.2	> 100
	Standards and statement I f	EN00210	Lagrag			
	Strychnos potatorum L.f.	EN00219	Leaves	DCM	0.9	60 100
		ENIODIOC		DCM/MeOH (1:1)	1.3	> 100
	Strychnos pungens Soler.	EN00186	Leaves	DCM	0.7	12.6
				DCM/MeOH (1:1)	0.8	80.4
				MeOH	0.8	80
				Water	2.3	39
hymelaeaceae	Gnidia cuneata Meisn.	EN00716	Leaves	DCM	0.7	31.1
nymenaeaeeae	Onulu cunculu Meisil.	LINOUVIO	Leaves	DCM/MeOH (1:1)	0.9	51.1
				Water	1.4	45.5
		EN100716	C +			
		EN00716	Stems	DCM	0.6	15.9
				DCM/MeOH (1:1)	0.8	40.5
	Gnidia kraussiana Meisn. var.	BP01008	Tuber	Water DCM/MeOH (1:1)	1.5 0.7	> 100 16
	kraussiana					
			.	Water	4.4	> 100
		BP01008	Leaves/twigs	DCM/MeOH (1:1)	0.4	10.8
				Water	1.3	> 100
iliaceae	Triumfetta welwitschii Mast. var. hirsuta (Sprague & Hutch.) Wild	BP01238	Leaves	DCM/MeOH (1:1)	1.0	3.6
	(Fright & Hatem) fild			Water	1.7	> 100
<i>V</i> erbenaceae	Clerodendrum glabrum E. Mey. var. glabrum	BP01207	Twigs	MeOH/DCM (1:1)	0.1	19
	č			Water	3.5	> 100
	Lantana camara L.	BP01216	Leaves/twigs	DCM/MeOH (1:1)	1.3	11

Table 1 (Continued)

Family	Plant species	Voucher number	Plant part	Solvent	% Yield	IC ₅₀ (µg/ml)
	<i>Lippia javanica</i> (Burm. f.) Spreng.	BP00200	Roots	DCM	0.8	3.8
				DCM/MeOH (1:1)	1.3	27
				MeOH	0.7	24
				Water	0.6	> 100
		BP00200	Stems	DCM	0.6	4.5
				DCM/MeOH (1:1)	1.7	21.8
				MeOH	0.8	29.8
				Water	1.0	> 100

(Celastraceae) (Brenniesen and Geisshusler, 1985; Carlini, 2003), *Conyza albida* Spreng. (Asteraceae) (Pacciaroni et al., 2000; Stamatis et al., 2003), *Lippia javanica* (Burm. f.) Spreng. (Verbenaceae) (Neidlein and Staehle, 1974; Van Wyk et al., 1997) and *Ocimum americanum* L. var. *americanum* (Lamiaceae) (Vieira et al., 2003), there are no reports on their antiplasmodial activity. Several representatives

of genera investigated here (viz. *Croton* (Euphorbiaceae) (Prozesky et al., 2001), *Acacia* (Fabaceae) (El Tahir et al., 1999b), *Hypericum* (Clusiaceae) (Decosterd et al., 1991; Gu et al., 1988), *Triumfetta* (Tiliaceae) (Muñoz et al., 2000) and *Vernonia* (Asteraceae) (Abosi and Raseroka, 2003; Kraft et al., 2003; Oketch-Rabah et al., 1998; Alves et al., 1997)) have previously been shown to have antiplasmodial

Table 2

Native or naturalised South African plants with high antiplasmodial activity, and comments on prior reports

	IC_{50} (µg/ml)	Comments			
Taxon					
Acacia tortilis	4.8	Acacia nilotica (L.) ex Delile has shown in vitro activity (El Tahir et al., 1999b)			
Aloe marlothii	3.5	In vitro activity previously reported (Van Zyl and Viljoen, 2002), although 16-fold lower than observed in this study			
Artemisia afra	5.0	In vitro activity previously reported; active constituents do not appear to be artemisinin compounds (Kraft et al., 2003)			
Bidens pilosa ^a 5.0		Watt and Breyer-Brandwijk (1962) reported negative antimalarial findings, but more recent test yielded positive in vitro results (Brandão et al., 1997; Krettli et al., 2001)			
Catha edulis 0.6		No previous reports of antiplasmodial activity			
Conyza albida ^a	2.0	No previous reports of antiplasmodial activity			
Croton gratissimus	3.5	Croton pseudopulchellus Pax has shown in vitro activity (Prozesky et al., 2001)			
Croton menyhartii	1.7	Croton pseudopulchellus Pax has shown in vitro activity (Prozesky et al., 2001)			
Euclea undulata	4.6	No previous reports of antiplasmodial activity			
Hypericum aethiopicum	1.4	<i>Hypericum japonicum</i> Thunb. (Gu et al., 1988) and <i>Hypericum calcyinum</i> L. (Decosterd et al., 1991) have shown in vivo and in vitro activity			
Kirkia wilmsii	3.7	No previous reports of antiplasmodial activity			
<i>Lippia javanica</i> 3.8		Reported to be a mosquito repellent (Govere et al., 2000), but no previous reports of antiplasmodial activity			
Ocimum americanum ^a	4.2	Used as a mosquito repellent (Seyoum et al., 2002), but no previous reports of antiplasmodial activity			
Pittosporum viridiflorum	3.0	No previous reports of antiplasmodial activity			
Plumbago zeylanica	3.0	In vitro activity previously reported (Simonsen et al., 2001)			
Ranunculus multifidus	2.3	No previous reports of antiplasmodial activity			
Schefflera umbellifera	3.7	In vitro activity previously reported (Tetyana et al., 2002)			
Species					
Setaria megaphylla	4.5	No previous reports of antiplasmodial activity			
Trichilia emetica	3.5	<i>Trichilia emetica</i> and other members of genus have shown in vitro activity (MacKinnon et al., 1997; El Tahir et al., 1999b; Prozesky et al., 2001)			
Triumfetta welwitschii	3.6	Triumfetta semitrilobata Jacq. has shown in vitro and in vivo activity (Muñoz et al., 2000)			
Vernonia colarata	4.7 Several <i>Vernonia</i> species have shown activity in vitro and in vivo (Alves et al., 1997; Oketch-Rabah et al., 1998; Abosi and Raseroka, 2003; Kraft et al., 2003)				
Vernonia myriantha	3.0	Several Vernonia species have shown activity in vitro and in vivo (Alves et al., 1997; Oketch-Rabah et al., 1998; Abosi and Raseroka, 2003; Kraft et al., 2003)			
Vernonia oligocephala	3.5	Several Vernonia species have shown activity in vitro and in vivo (Alves et al., 1997; Oketch-Rabah et al., 1998; Abosi and Raseroka, 2003; Kraft et al., 2003)			

^a Naturalised in South Africa.

activity. In addition to identifying further species within these genera that display activity, our findings are substantiated by earlier reported antiplasmodial activity elsewhere in the genera. Similarly, the results for Plumbago zeylanica L. (Plumbaginaceae) (Simonsen et al., 2001), Bidens pilosa L. (Asteraceae) (Brandão et al., 1997; Krettli et al., 2001), Trichilia emetica Vahl (Meliaceae) (El Tahir et al., 1999a; Prozesky et al., 2001), Schefflera umbellifera (Sond.) Baill. (Araliaceae) (Tetyana et al., 2002) and Artemisia afra Jacq. ex Willd. (Asteraceae) (Kraft et al., 2003) agree with previous reports on their antiplasmodial activity. The activity of Aloe marlothii A. Berger (Asphodelaceae) has been reported on before (IC₅₀ > 50 μ g/ml), although it was considerably lower than that observed in this study (Van Zyl and Viljoen, 2002). A number of the genera investigated, namely, Euclea (Ebenaceae), Kirkia (Kirkiaceae), Pittosporum (Pittosporaceae), Ranunculus (Ranunculaceae) and Setaria (Poaceae) were shown for the first time to display high antiplasmodial activity. To the best of our knowledge, this is the first report of the antiplasmodial activity in the families Ebenaceae, Kirkiaceae and Pittosporaceae.

It is interesting to note that the majority of the aqueous extracts, which would be the preferred method of preparing the plants when used in traditional medicines, did not show any activity. Moreover, they would contain very little if any of the lipophilic compounds extracted with the organic solvents (DCM and MeOH), which showed the greatest activity. A possible explanation for the poor hit rate of aqueous extracts is that they were not prepared according to the traditional methods, which often involves boiling for several hours. Furthermore, in the traditional context it is not uncommon for several plant taxa to be administered as mixtures. In this case, synergism could exist between the various phytochemicals or different constituents could help extract and keep active lipophilic compounds in an aqueous solution. In the course of this study the above-mentioned factors were not taken into consideration and only the in vitro antiplasmodial activity of individual plant extracts was determined.

A number of the plants selected did not display in vitro antiplasmodial activity, despite strong associations with malaria and its treatment. A possible explanation could be that the plants act as antipyretics or immune stimulants to relieve the symptoms of the disease, rather than having direct antiparasitic activity (Phillipson et al., 1993). Alternatively, precursors of the active components may be present in the extracts but have to be modified, usually in vivo, before activity is exhibited. Furthermore, factors such as chemotypes, environmental parameters, harvesting and storage conditions could collectively influence the plant secondary metabolites prior to and following harvesting, which in turn would be reflected in the bioactivity. Despite these possible sources of variability, a number of the plant species that were found to be active in this study have previously been shown to possess in vitro, and in some cases in vivo, antiplasmodial activity. In the course of this study, several taxa at the genus and family levels were shown for the first time to display in vitro antiplasmodial activity and warrant further investigation as potential sources of antiplasmodial agents. Additional in vitro and in vivo work aimed at understanding the mechanisms of action of the active plant species and isolating and characterising the bioactive constituents is underway in our laboratories and will be reported on in due course.

4. Conclusions

Not only has this study highlighted promising taxa for further antimalarial investigation, it has provided compelling evidence for the rational exploration of indigenous and naturalised South African medicinal plants as a source of antiplasmodial agents. Considering that most regional plant taxa have not been investigated chemically or pharmacologically, they remain a potential source of leads for drug development. This is particularly important for diseases lacking effective chemotherapeutic agents, such as malaria.

Acknowledgements

We gratefully acknowledge the Department of Science and Technology of South Africa for the Innovation Fund Grant (Project 31313). We wish to acknowledge the traditional knowledge holders of South Africa for their contribution to the indigenous knowledge systems of South Africa, and the many genetic resource holders who allowed access to their plant materials. In particular, N. Hahn and E. Schmidt are thanked for facilitating collections on their properties. In this regard, we also thank South Africa's provincial conservation authorities for permitting plant collections. We wish to thank the research and field teams of participating institutions. Miss S. Luwaca is thanked for administrative support.

References

- Abosi, A.O., Raseroka, B.H., 2003. In vivo antimalarial activity of Vernonia amygdalina. British Medical Journal of Biomedical Science 60, 89–91.
- Alves, T.M., Nagem, T.J., de Carvalho, L.H., Krettli, A.U., Zani, C.L., 1997. Antiplasmodial triterpenes from *Vernonia brasiliana*. Planta Medica 63, 554–555.
- Arnold, T.H., Prentice, C.A., Hawker, L.C., Snyman, E.E., Tomalin, M., Crouch, N.R., Pottas-Bircher, C., 2002. Medicinal and Magical Plants of Southern Africa: An Annotated Checklist. Strelitzia 13. National Botanical Institute, Pretoria, pp. 1–2.
- Brandão, M.G.L., Krettli, A.U., Soares, L.S.R., Nery, C.G.C., Marinuzzi, H.C., 1997. Antimalarial activity of extracts and fractions from *Bidens pilosa* and *Bidens* species (Asteraceae) correlated with the presence of acetylene and flavonoid compounds. Journal of Ethnopharmacology 57, 131–138.
- Brenniesen, R., Geisshusler, S., 1985. Psychotropic drugs III: analytical and chemical aspects of *Catha edulis* Forsk. Pharmacetica Acta Halvetiae 60, 290–301.

- Carlini, E.A., 2003. Plants and the central nervous system. Pharmacology, Biochemistry and Behavior 75, 501–512.
- Clark, T.E., Appleton, C.C., Drewes, S.E., 1997. A semi-quantitative approach to the selection of appropriate candidate molluscicides—a South African application. Journal of Ethnopharmacology 56, 1–13.
- Clarkson, C., Campbell, W.E., Smith, P., 2003. In vitro antiplasmodial activity of abietane and totarane diterpenes isolated from *Harpagophytum procumbens* (Devil's Claw). Planta Medica 8, 720–724.
- Decosterd, L.A., Hoffmann, E., Kyburz, R., Bray, D., Hostettmann, K., 1991. A new phloroglucinol derivative from *Hypericum calcyinum* with antifungal and in vitro antimalarial activity. Planta Medica 57, 548–551.
- El Tahir, A., Satti, G.M., Khalid, S.A., 1999a. Antiplasmodial activity of selected Sudanese medicinal plants with emphasis on *Maytenus* senegalensis (Lam.) Exell. Journal of Ethnopharmacology 64, 227–233.
- El Tahir, A., Satti, G.M., Khalid, S.A., 1999b. Antiplasmodial activity of selected Sudanese medicinal plants with emphasis on *Acacia nilotica*. Phytotherapy Research 13, 474–478.
- Govere, J., Durrheim, D.N., Du Toit, N., Hunt, R.H., Coetzee, M., 2000. Local plants as repellents against *Anopheles arabiensis*, in Mpumalanga Province, South Africa. Central African Journal of Medicine 46, 213– 216.
- Gu, G., Feng, S., Xiaoyan, W., 1988. Antimalarial constituents of *Hyper-icum japonicum* Thunb. Isolation and structure of japonicins A, B, C and D. Huaxue Xuebao 46, 246–251.
- Kraft, C., Jenett-Siems, K., Siems, K., Jakupovic, J., Mavi, S., Bienzle, U., Eich, E., 2003. In vitro antiplasmodial evaluation of medicinal plants from Zimbabwe. Phytotherapy Research 17, 123–128.
- Krettli, A.U., Andrade-Neto, V.F., Brandão, M.G.L., Ferrari, W.M.S., 2001. The search for new antimalarial drugs from plants used to treat fever and malaria or plants randomly selected: a review. Memorias do Instituto Oswaldo Cruz 96, 1033–1042.
- MacKinnon, S., Durst, T., Arnason, J.T., Angerhofer, C., Pezzuto, J., Sanchez-Vindas, P.E., Poveda, L.J., Gbeassor, M., 1997. Antimalarial activity of tropical Meliaceae extracts and Gedunin derivatives. Journal of Natural Products 60, 336–341.
- Makler, M.T., Ries, J.M., Williams, J.A., Bancroft, J.E., Piper, R.C., Gibbins, B.L., et al., 1995. Parasite lactate dehydrogenase as an assay for *Plasmodium falciparum* drug sensitivity. American Journal of Tropical Medicine and Hygiene 48, 739–741.
- Muñoz, V., Sauvain, M., Bourdy, G., Callapa, J., Rojas, I., Vargas, L., Tae, A., Deharo, E., 2000. The search for natural bioactive compounds through a multidisciplinary approach in Bolivia. Part II. Antimalarial activity of some plants used by Mosetene Indians. Journal of Ethnopharmacology 69, 139–155.
- Neidlein, R., Staehle, R., 1974. Constituents of *Lippia javanica*. III. Deutsche Apotheker Zeitung 114, 1588–1592.
- Oketch-Rabah, H.A., Christensen, S.B., Frydenvang, K., Dossaji, S.F., Theander, T.G., Cornett, C., Watkins, W.M., Kharazmi, A., Lemmich,

E., 1998. Antiprotozoal properties of 16,17-dihydrobrachycalyxolide from *Vernonia brachycalyx*. Planta Medica 64, 559–562.

- Pacciaroni, A.V., Mongelli, E., Ariza Espinar, L., Romano, A., Ciccia, G., Silva, G.L., 2000. Bioactive constituents of *Conyza albida*. Planta Medica 66, 720–723.
- Phillipson, J.D., Wright, C.W., Kirby, G.C., Warhurst, D.C., 1993. Tropical plants as sources of antiprotozoal agents. Recent Advances in Phytochemistry 27, 1–40.
- Prozesky, E.A., Meyer, J.J.M., Louw, A.I., 2001. In vitro antiplasmodial activity and cytotoxicity of ethnobotanically selected South African plants. Journal of Ethnopharmacology 76, 239–245.
- Ridley, R.G., 2002. Medical need, scientific opportunity and the drive for antimalarial drugs. Nature 415, 686–693.
- Seyoum, A., Palsson, K., Kung'a, S., Kabiru, E.W., Lwande, W., Killeen, G.F., Hassanali, A., Knols, B.G.J., 2002. Traditional use of mosquito-repellent plants in western Kenya and their evaluation in semi-field experimental huts against *Anopheles gambiae*: ethnobotanical studies and application by thermal expulsion and direct burning. Transactions of the Royal Society of Medicine and Hygiene 96, 225– 231.
- Simonsen, H.T., Nordskjold, J.B., Smitt, U.W., Nyman, U., Palpu, P., Joshi, P., Varughese, G., 2001. In vitro screening of Indian medicinal plants for antiplasmodial activity. Journal of Ethnopharmacology 74, 195–204.
- Stamatis, G., Kyriazopoulos, P., Golegou, S., Basayiannis, A., Skaltsas, S., Skaltsa, H., 2003. In vitro anti-*Helicobacter pylori* activity of Greek herbal medicines. Journal of Ethnopharmacology 88, 175–179.
- Tetyana, P., Prozesky, E.A., Jäger, A.K., Meyer, J.J.M., van Staden, J., 2002. Some medicinal properties of *Cussonia* and *Schefflera* species used in traditional medicine. South African Journal of Botany 68, 51– 54.
- Trager, W., Jensen, J.B., 1976. Human malaria parasites in continuous culture. Science 193, 673–675.
- Van Wyk, B.E., 2002. A review of ethnobotanical research in southern Africa. South African Journal of Botany 68, 1–13.
- Van Wyk, B.E., van Oudtshoorn, B., Gericke, N., 1997. Medicinal Plants of South Africa. Briza Publications, Pretoria, South Africa.
- Van Zyl, R.L., Viljoen, A.M., 2002. In vitro activity of *Aloe* extracts against *Plasmodium falciparum*. South African Journal of Botany 68, 106–110.
- Vieira, R.F., Grayer, R.J., Paton, A.J., 2003. Chemical profiling of *Ocimum americanum* using external flavonoids. Phytochemistry 63, 555– 567.
- Watt, J.M., Breyer-Brandwijk, M.G., 1962. The Medicinal and Poisonous Plants of Southern and Eastern Africa. E. & S. Livingstone Ltd., Edinburgh, UK.
- WHO, 2003. World Health Organisation Fact Sheet No. 94. WHO information. http://www.int/inf~fs/en/fact094.html.